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# Complex of C<sub>60</sub> Fullerene with Doxorubicin as a Promising Agent in Antitumor Therapy

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## Abstract

The main aim of this work was to evaluate the effect of doxorubicin in complex with C<sub>60</sub> fullerene (C<sub>60</sub> + Dox) on the growth and metastasis of Lewis lung carcinoma in mice and to perform a primary screening of the potential mechanisms of C<sub>60</sub> + Dox complex action. We found that volume of tumor from mice treated with the C<sub>60</sub> + Dox complex was 1.4 times less than that in *control* untreated animals. The number of metastatic foci in lungs of animals treated with C<sub>60</sub> + Dox complex was two times less than that in *control* untreated animals. Western blot analysis of tumor lysates revealed a significant decrease in the level of heat-shock protein 70 in animals treated with C<sub>60</sub> + Dox complex. Moreover, the treatment of tumor-bearing mice was accompanied by the increase of cytotoxic activity of immune cells. Thus, the potential mechanisms of antitumor effect of C<sub>60</sub> + Dox complex include both its direct action on tumor cells by inducing cell death and increasing of stress sensitivity and an immunomodulating effect. The obtained results provide a scientific basis for further application of C<sub>60</sub> + Dox nanocomplexes as treatment agents in cancer chemotherapy.

**Keywords:** C<sub>60</sub> fullerene, Doxorubicin, Antitumor effect, Antimetastatic effect, Immune response

## Background

Suppression of proliferative activity of tumor cells is a basic strategy when using traditional chemotherapeutic drugs [1]. Doxorubicin (Dox) is the anthracycline antibiotic widely used for treatment of cancers of different origin [2]. It uses two main mechanisms in cytotoxic action towards tumor cells: intercalation into DNA helices followed by inhibition of the DNA synthesis and generation of free radicals followed by DNA impairment and cell membrane damage [3]. However, the antitumor effect of traditional chemotherapy is always associated with numerous negative side effects, in particular, the toxicity towards cells of normal organs and tissues. Dox causes potent oxidative stress, mitochondrial dysfunction, and Bcl-2 expression disturbance followed by the apoptotic damage in heart tissue. A number of substances with ability to attenuate the Dox-induced cardiotoxicity have been developed nowadays in order to improve the outcome of the long-term treatment with

Dox [4]. In that regard, C<sub>60</sub> fullerene is a promising carbon nanostructure that is characterized by unique physical and chemical properties [5] and biological activity both *in vitro* and *in vivo* [6].

It was established that pristine C<sub>60</sub> fullerenes at low concentrations are nontoxic [7, 8] and they are able to penetrate through the cytoplasmic membrane of treated cells [9]. One of the biologically most relevant features of C<sub>60</sub> fullerene is its antioxidant effect [10]. Our previous results also revealed antioxidant properties of pristine C<sub>60</sub> fullerene [11].

C<sub>60</sub> fullerene and its derivatives possess potent anticancer activity [12]. It has been reported that C<sub>60</sub> fullerene nanocrystal induces certain hallmarks of autophagy in cancer cells [13]. The tumor inhibitory effect of fullerenes is accompanied by the immunomodulatory activity [14].

It is important to emphasize that chemical modification of the surface of C<sub>60</sub> molecule for improvement of its water solubility often leads to changes in its physical and chemical properties and to a decrease in specific biological effects. Thus, utilization of pristine C<sub>60</sub> fullerene would be more desirable. In our previous study [15], we showed that the water-soluble pristine C<sub>60</sub> fullerene

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directly suppresses growth of transplanted malignant tumor. Chen et al. [16] reported that antitumor effect of pristine and functionalized C<sub>60</sub> fullerene might be associated with modulation of the oxidative stress and the anti-angiogenic and immunostimulatory activity. Injac et al. [17] demonstrated the ability of fullerenol to decrease the acute Dox pulmototoxicity in rats with malignant neoplasm through inhibition of oxidative stress. In our previous study [18], it was shown that the combination of Dox with C<sub>60</sub> fullerene resulted in increase of therapeutic efficacy of the treatment. Taking into account these data, we also suggested [19, 20] that Dox immobilization on C<sub>60</sub> fullerene (C<sub>60</sub> + Dox complex formation) can reduce negative side effects of this drug towards normal cells as well as enhance its ability to enter target tumor cells.

The main goal of this work was to (1) evaluate the effect of C<sub>60</sub> + Dox complex on growth and metastasis of Lewis lung carcinoma (LLC) and (2) perform primary screening of the potential mechanisms of C<sub>60</sub> + Dox action.

## Methods

### Material Preparation and Characterization

The highly stable aqueous colloid solution of purified C<sub>60</sub> fullerene (C<sub>60</sub>FAS; concentration 0.15 mg/ml) was prepared as reviewed in [21, 22]. The method is based on the technology of transferring C<sub>60</sub> molecules from toluene to an aqueous phase with the help of ultrasonic treatment.

The atomic force microscopy (AFM) data [21–23] confirm randomly arranged individual C<sub>60</sub> molecules with a diameter of ~0.7 nm and their bulk sphere-like aggregates with a height of 2–100 nm in C<sub>60</sub>FAS.

Dox (“Doxorubicin-TEVA”, Pharmachemie B.V.) was dissolved in saline at initial concentration 0.15 mg/ml and used in all experiments. It was immobilized on the C<sub>60</sub> fullerene according to the following protocol: C<sub>60</sub>FAS (0.15 mg/ml) and Dox (0.15 mg/ml) were mixed in 1:2 volume ratio, and the resulting mixture was treated for 20 min in the ultrasonic disperser. After that, it was subjected to 12-h magnetic stirring at room temperature. Pronounced hypochromic effect observed in spectrophotometric experiment and AFM data clearly indicate a formation of stable C<sub>60</sub> + Dox complex [19, 20].

### Animals

The male C57Bl/6 mice (20–21 g weight) were kept at 298 ± 1 K on a standard diet in the vivarium of R.E. Kavetsky Institute of Experimental Pathology, Oncology and Radiobiology, NAS of Ukraine (Kyiv). All experiments were conducted in accordance with the international principles of the European Convention for

protection of vertebrate animals under the control of Bio-Ethics Committee of that institution.

### Tumor Model, Treatment Regimens, and Study Design

LLC was used as an experimental model. LLC cell line was obtained from the cell line bank of the R.E. Kavetsky Institute of Experimental Pathology, Oncology and Radiobiology, NAS of Ukraine (Kyiv). Tumor cells ( $5 \times 10^5$  in the volume of 100 μl) were transplanted intramuscularly into the mouse limb. After transplantation of tumor cells, the experimental animals were randomized by weight and distributed in four groups with ten animals per group:

*Group 1* (C<sub>60</sub> fullerene injection). C<sub>60</sub>FAS was used in 1.5 mg/kg dose (0.2 ml) injected intraperitoneally to mice with transplanted tumor once per day for 5 days with a day interval [15].

*Group 2* (Dox injection). Dox was used in 1.5-mg/kg dose (0.2 ml) injected intraperitoneally to mice with transplanted tumor once per day for 5 days with a day interval [24].

*Group 3* (C<sub>60</sub> + Dox complex injection). C<sub>60</sub> + Dox mixture was used in 1.5-mg/kg dose (0.2 ml) injected intraperitoneally to mice with transplanted tumor once per day for 5 days with a day interval.

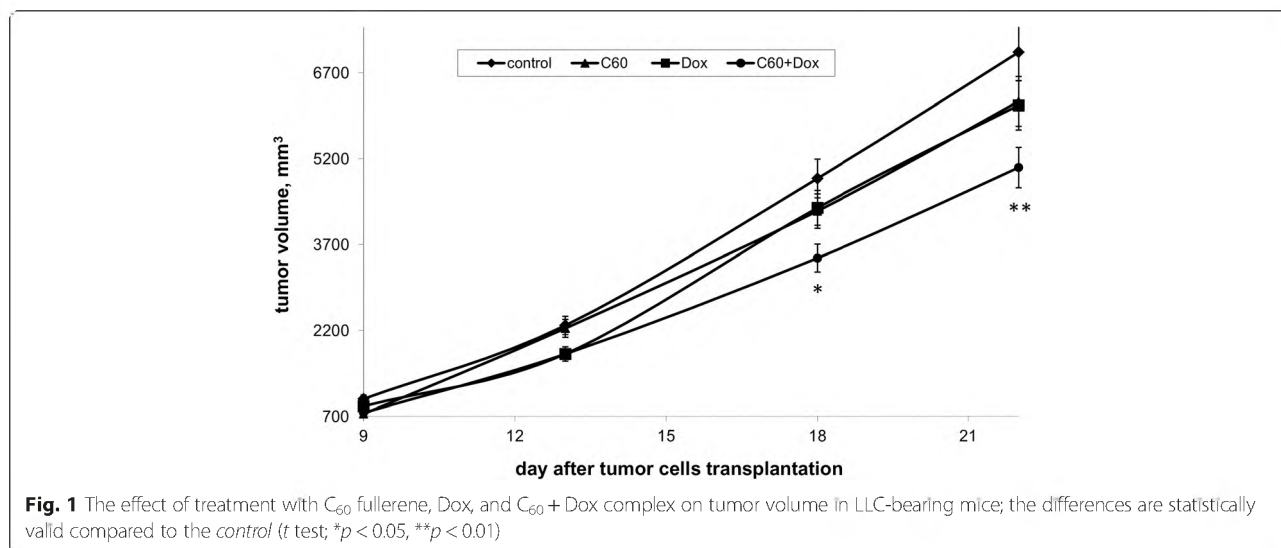
*Control*. The mice with transplanted tumor were injected with saline (0.2 ml) once per day for 5 days with a day interval.

*Intact animals* were used in order to investigate immunological indices (cytotoxic activity of the peritoneal macrophages and mononuclear splenic leucocytes).

The injections of C<sub>60</sub> fullerene, Dox, or C<sub>60</sub> + Dox complex were started on the 2nd day after tumor cell transplantation. The protocol of injecting C<sub>60</sub> fullerenes was based on the fact that C<sub>60</sub> fullerenes administered intraperitoneally to rats (500 mg/kg) were subjected to clearance from the organism within 2–4 days [25]. The C<sub>60</sub> fullerene dose applied in our experiments was significantly lower than the LD<sub>50</sub> value determined for C<sub>60</sub> fullerene which, after oral administration to mice, was equivalent to 600 mg/kg of body weight [25].

The kinetics of tumor growth was evaluated as described [15] by linear dimensions of tumor measured every third day with the use of calipers starting from the 9th day after tumor cell inoculation. The euthanasia of experimental animals was performed at the end of the experiment (22nd day), and the number and size of metastases in animal lungs were monitored.

Anticancer effect was also characterized by growth inhibition index, GII, calculated by the formula  $GII = (V_c - V_{exp})/V_c \times 100 \%$ , where  $V_c$  and  $V_{exp}$  are the average volumes of tumor of control and experimental



animals, respectively;  $V = 1/2 \cdot \left(\frac{a+b}{2}\right)^3$ , where  $a$  and  $b$  are the length and width (in millimeters) of the tumor site, respectively [15].

**MTT Assay**

To analyze cytotoxic activity of the peritoneal macrophages and mononuclear splenic leukocytes, the modified MTT assay was used as described [26]. Cytotoxic activity of the studied samples was calculated using the formula Cytotoxicity index =  $(1 - \epsilon/\epsilon_c) \times 100\%$ , where  $\epsilon_c$  and  $\epsilon$  are the extinctions of control and test sample, respectively. Measurement of extinction was performed on a digital spectrophotometer ( $\mu$ Quant, BioTEK, USA) at the wavelength of 540 nm.

The investigation of cytotoxic activity of immunocytes was performed on the 22nd day after tumor cell transplantation. Suspension of tumor cells was prepared from tissue homogenates. Mononuclear splenic leukocytes were obtained from splenocyte suspension by centrifugation (1500 rpm, 40 min) in Ficoll-Hypaque density gradient ( $p = 1.077$ ). Peritoneal macrophages were isolated without preliminary stimulation. Mice were sacrificed, and peritoneal macrophages were harvested using phosphate-buffered saline containing 100 U/ml of heparin. Cells were centrifuged at 300xg for 5 min at 4 °C, washed twice with serum-free DMEM, and re-suspended in DMEM containing 10 % FCS and 40  $\mu$ g/ml gentamicin.

To perform cytotoxic assay, LLC cells were placed in 96-well plates ( $3 \times 10^5$  cells/well), and mononuclear splenic leukocytes or peritoneal macrophages were added at 20:1 ratio. Cells were incubated in a RPMI-1640 medium supplemented with gentamicin sulfate (100  $\mu$ g/ml) and maintained at 37 °C for 18 h in 5 % CO<sub>2</sub> atmosphere. After incubation, MTT (Sigma) was added to a final

concentration of 0.5 mg/ml followed by culturing for 3 h. After culturing, cells were centrifuged at 4000 rpm (1600xg) for 10 min. Culture medium was removed, and blue formazan crystals were dissolved in 100  $\mu$ l DMSO. Optical density was determined at 570 nm.

**Western Blot Analysis**

The tumors were surgically removed, and cell lysates were prepared by EDTA extraction. After removal of unlysed cell remnants and nuclei by centrifugation in the Eppendorf micro-centrifuge (5 min, 10,000 rpm, 10,200xg); protein concentration was determined by standard method, as described [27]; and 10  $\mu$ g of equal amounts of protein was loaded into 15 % polyacrylamide gel. Proteins were resolved and transferred to Immobilon-P membrane (Millipore, Billerica, MA) using semi-dry transfer (Bio-Rad, Hercules, CA). After incubating the membrane in the blocking buffer, the membrane was incubated with heat-shock protein 70 (HSP70) monoclonal antibodies (Santa Cruz Biotechnology, Santa Cruz, CA). For a loading control, the levels of expression of the  $\beta$ -actin were detected in each sample using mouse  $\beta$ -actin monoclonal antibodies (Sigma). Immunoreactive bands were visualized by chemiluminescence using horseradish

**Table 1** Tumor growth inhibition index (GIi, %) for each experimental group on the 9th, 13th, 18th, and 22nd day after tumor inoculation

Animal group	Day after tumor cell transplantation			
	9	13	18	22
Group 1 (C <sub>60</sub> fullerene injection), n = 10	26	20	22	12
Group 2 (Dox injection), n = 10	13	22	21	13
Group 3 (C <sub>60</sub> + Dox injection), n = 10	24	22	29	29

**Table 2** The effect of C<sub>60</sub> fullerene and Dox used alone and in C<sub>60</sub> + Dox complex on the LLC metastases

Animal group	The number of tumor nodules in lungs							Total number
	Nodule diameter (mm)							
	<0.5	0.5	1	2	3	4	5	
Control, n = 10	12	9	33	25	3	7	11	100
Group 1(C <sub>60</sub> fullerene injection), n = 10	20	22	23	8	9	1	1	84
Group 2(Dox injection), n = 10	14	5	25	16	7	1	1	69
Group 3(C <sub>60</sub> + Dox injection), n = 10	16	10	6	16	–	–	–	48

peroxidase-conjugated IgG antibodies and ECL Kit (Amersham, Uppsala, Sweden) according to the instructions of the manufacturer.

**Statistical Analysis**

For statistical analysis of the obtained results, standard variation data within a group was calculated together with a statistical reliability of differences between two groups of data assessed by Student’s *t* test. The level of significance was set to *p* < 0.05.

**Results and Discussion**

**Treatment with C<sub>60</sub> + Dox Complex Results in the Inhibition of Tumor Growth and Metastasis and Increases Stress Sensitivity of Tumor Cells In Vivo**

Experimental animals tolerated the treatment well and exhibited normal behavior, as determined by the activity level and grooming behavior throughout the study.

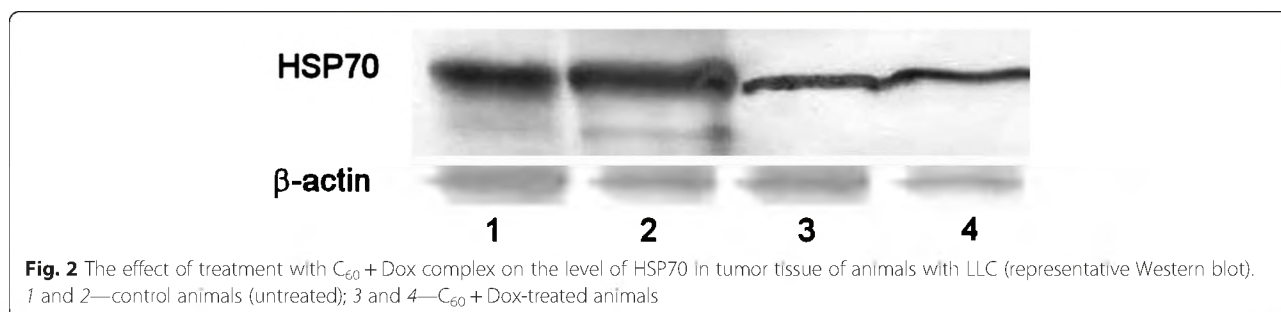
LLC was characterized by a significant growth of its size (volume) from the 9th to the 22nd day of the experiment (Fig. 1). One can see that all applied treatments (C<sub>60</sub> fullerene, Dox, and C<sub>60</sub> + Dox complex) caused a decrease (comparing to *control*, i.e., untreated mice) of tumor volume. Tumor volume in animals of *group 1* (C<sub>60</sub> fullerene injection) and *group 2* (Dox injection) differed slightly. The volume of tumor from mice treated with the C<sub>60</sub> + Dox complex was significantly lower than that in *control* untreated animals, viz. by 1.4 times.

The effect of C<sub>60</sub> fullerene and Dox used alone and in C<sub>60</sub> + Dox complex on tumor growth was evaluated by the GII value presented in Table 1.

Thus, on the 13th day after cancer cell inoculation, tumor volume in mice treated with Dox and C<sub>60</sub> + Dox

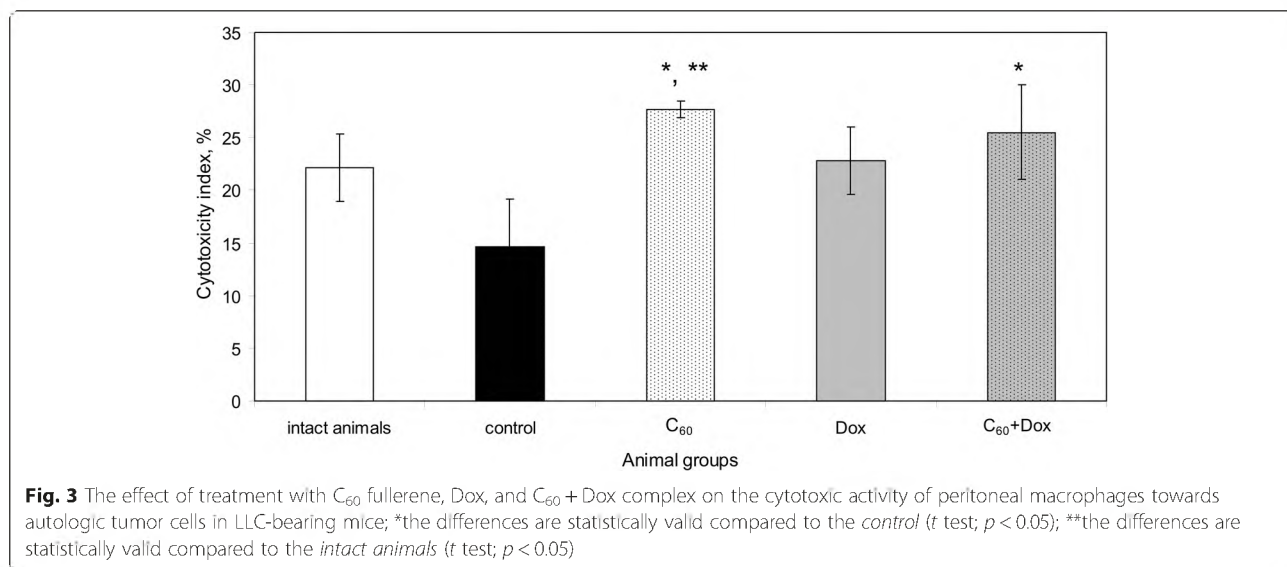
complex was 22 % less than that in the untreated animals. There was no inhibition of tumor growth in mice treated with C<sub>60</sub> fullerene alone. On the 18th day after cancer cell transplantation, we observed the most expressed retardation of tumor growth in animals treated with C<sub>60</sub> + Dox complex, and the GII value in mice of that group was 29 %. At that time point, the tumor volume in mice treated with C<sub>60</sub> fullerene and Dox was ~22 % less than that in the untreated animals. On the 22nd day after cancer cell inoculation, the GII value in mice treated with C<sub>60</sub> fullerene and Dox was decreased by ~13 %, but it did not change in mice treated with C<sub>60</sub> + Dox complex.

The treatment of tumor-bearing mice with C<sub>60</sub> fullerene, Dox, and C<sub>60</sub> + Dox complex caused an inhibition of metastasis of the experimental tumor (Table 2). The number of the metastatic foci in lungs of animals of the group treated with C<sub>60</sub> + Dox complex was two times less than that in the untreated animals and 1.4 times less than that in mice treated with Dox alone. It should be noted that the metastatic foci in mice treated with C<sub>60</sub> fullerene, Dox, and its complex were characterized by different sizes. While in the *control* group, large metastatic foci that infiltrated into the lung parenchyma were observed; the metastatic foci were much smaller and solitary in mice treated with Dox and C<sub>60</sub> + Dox complex. In mice treated with C<sub>60</sub> + Dox complex, the metastatic foci with the diameter of ≥3 mm were absent. Since only tumor growth beyond the size of 1–2 mm is angiogenesis-dependent [28], we suggested that the small-sized metastatic focus (≤1 mm in diameter) is in a state of dormancy. Therefore, one can suppose that C<sub>60</sub> + Dox complex exerts a negative effect towards tumor angiogenesis.



**Fig. 2** The effect of treatment with C<sub>60</sub> + Dox complex on the level of HSP70 in tumor tissue of animals with LLC (representative Western blot). 1 and 2—control animals (untreated); 3 and 4—C<sub>60</sub> + Dox-treated animals



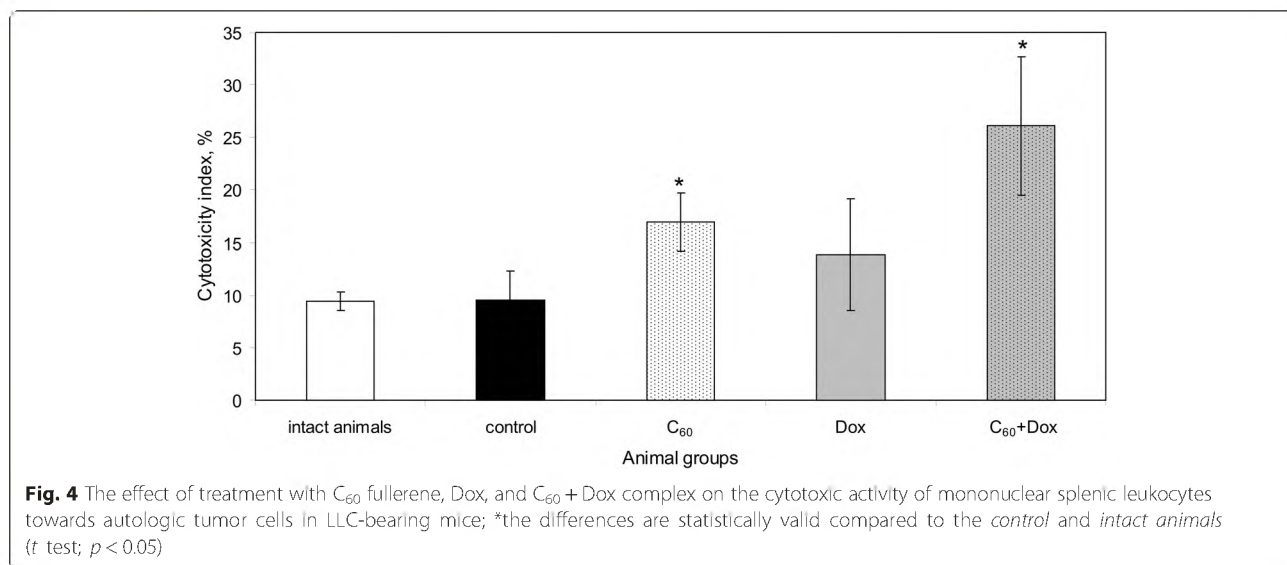


It is known that HSP70 is aberrantly expressed in cancer cells of different origins. The survival of these cells strongly depends upon HSPs due to their role not only in protein refolding and degradation but also in preventing apoptosis [29]. Elevated expression of HSP70 is associated with poor response of tumor cells to chemotherapy, and inhibition of its expression was shown to be an effective strategy against cancer [30]. Therefore, we have measured the level of HSP70 in tumor tissue of animals treated with Dox and C<sub>60</sub> + Dox complex. Western blot analysis of tumor cell lysates revealed a significant decrease in HSP70 level only in animals treated with C<sub>60</sub> + Dox complex (Fig. 2). The level of HSP70 in tumor lysates from mice treated with Dox alone did not differ from that in the untreated tumor-bearing mice.

The obtained results clearly demonstrate that the anti-cancer activity of Dox is not only well preserved in its complex with C<sub>60</sub> fullerene, but it is even enhanced after formation of such complex.

**C<sub>60</sub> + Dox Complex Modulates Immunological Reactivity of Tumor-Bearing Mice**

C<sub>60</sub> fullerene and its derivatives were shown to possess the immunomodulating properties [14]. Thus, we supposed that the immunomodulating effect of C<sub>60</sub> + Dox complex can be involved in its antitumor action. To testify this hypothesis, the cytotoxic activity of mononuclear splenic leukocytes and macrophages towards autologous tumor cells was evaluated in tumor-bearing mice treated with the C<sub>60</sub> + Dox complex.



Growth of experimental tumor was associated with a decrease of macrophage cytotoxicity towards autologous tumor cells *in vitro* (Fig. 3).

Treatment with Dox as well as with C<sub>60</sub> fullerene used alone and in C<sub>60</sub> + Dox complex resulted in increased cytotoxic activity of the peritoneal macrophages of tumor-bearing mice. Cytotoxic indices of the peritoneal phagocytes in treated animals were comparable with those in the intact mice.

Cytotoxic activity of splenic mononuclear leukocytes in tumor-bearing mice did not differ from that in the intact animals (Fig. 4).

Treatment with Dox resulted in an increase of splenocyte cytotoxicity. A significant individual variability of cytotoxic indices in animals from this group should be noted. Cytotoxicity indices of the mononuclear splenic leukocytes in animals treated with C<sub>60</sub> fullerene used alone and in C<sub>60</sub> + Dox complex were significantly higher than those in the untreated tumor-bearing mice. The most positive effect was observed in animals receiving C<sub>60</sub> + Dox complex. Cytotoxic activity of splenic mononuclear cells towards autologous tumor cells is substantially mediated by splenic NK cells [31]. Turabekova M et al. reported that C<sub>60</sub> fullerene might be recognized by Toll-like receptors (TLRs) [32]. TLR-dependent stimulatory effect of the preparation and an increased stress sensitivity of LLC cells associated with a decreased HSP70 expression might be potential reasons of increased cytotoxicity of splenic mononuclear cells in animals receiving C<sub>60</sub> + Dox complex.

## Conclusions

The results of performed experiments demonstrated that treatment of mice bearing LLC with C<sub>60</sub> + Dox nanocomplexes is associated with a significant antitumor effect, namely, (1) the volume of tumor of mice treated with C<sub>60</sub> + Dox complex was 1.4 times smaller than that in the *control* untreated animals; (2) the number of metastatic foci in lungs of animals of the group treated with C<sub>60</sub> + Dox complex was two times smaller than that in *control* untreated animals; (3) there were no metastatic foci with diameter  $\geq 3$  mm in mice treated with C<sub>60</sub> + Dox complex. Western blot analysis of tumor cell lysates of animals treated with C<sub>60</sub> + Dox complex revealed a significant decrease in the HSP70 level. The MTT assay showed that C<sub>60</sub> + Dox complex modulates immunological reactivity of tumor-bearing mice. The potential mechanisms of C<sub>60</sub> + Dox complex antitumor effect are likely to be based on its direct action on tumor cells with inducing cell death as well as an increasing of stress sensitivity and immunomodulating effect. Thus, the C<sub>60</sub> + Dox nanocomplexes might be proposed as new pharmacological agents that are effectively killing tumor cells and simultaneously stimulating immune responses in tumor-bearing mice.

## Competing Interests

The authors declare that they have no competing interests.

## Authors' Contributions

RS and UR designed the experiments; YP, ME, and PS were involved in the synthesis and characterization of the samples; SP and RP have done *in vitro* studies of C<sub>60</sub>-Dox complexes (MTT assay, Western blot analysis); and SP, LS, and GD were responsible for *in vivo* studies of C<sub>60</sub>-Dox complexes. GP, UR, and SP analyzed the data of experiments. SP, YP, and UR wrote the manuscript of the paper. All authors read and approved the final manuscript.

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## References

- Hirsch J (2006) *JAMA* 296:1518
- Abou EHMA, Verheul HM, Jorna AS, Schalkwijk C, van Bezu J, van der Vijgh WJ, Bast A (2003) *Brit J Cancer* 98:357
- Szulawska A, Czyz M (2006) *Postepy Hig Med Dosw* 60:78
- Jay SM, Murthy AC, Hawkins JF, Wortzel JR, Steinhilber ML, Alvarez LM, Gannon J, Macrae CA, Griffith LG, Lee RT (2013) *Circulation* 128:152
- Fullerenes: Principles and Applications (2nd Ed) (2011). In: Langa De La Puente F, Nierengarten J-F (eds). Cambridge: The Royal Society of Chemistry. p 650. doi:10.1039/9781849732956-PP001.
- Anilkumar P, Lu F, Cao L, Luo P, Liu JH, Sahu S, Tackett K II, Wang Y, Sun YP (2011) *Current Med Chem* 18:2045
- Prylutska SV, Matyshevska OP, Golub AA, Prylutskyi YI, Potebnyia GP, Ritter U, Scharff P (2007) Study of C<sub>60</sub> fullerenes and C<sub>60</sub>-containing composites cytotoxicity *in vitro*. *Mater Sci Engineer C* 27:1121–1124
- Johnston HJ, Hutchison GR, Christensen FM, Aschberger K, Stone V (2010) *Toxicol Sci* 114:162
- Prylutska S, Bilyi R, Overchuk M, Bychko A, Andreichenko K, Stoika R, Rybalchenko V, Prylutskyi Y, Tsierekzos NG, Ritter U (2012) Water-soluble pristine fullerenes C<sub>60</sub> increase the specific conductivity and capacity of lipid model membrane and form the channels in cellular plasma membrane. *J Biomed Nanotechnol* 8:522–527
- Markovic Z, Trajkovic V (2008) *Biomaterials* 29:3561
- Prylutska SV, Grynyuk II, Matyshevska OP, Prylutskyi YI, Ritter U, Scharff P (2008) Anti-oxidant properties of C<sub>60</sub> fullerenes *in vitro*. *Fullerenes, Nanotubes, Carbon Nanostruct* 16:698–705
- Chen Z, Mao R, Liu Y (2012) *Curr Drug Metabolism* 13:1035
- Zhang Q, Yang W, Man N, Zheng F, Shen Y, Sun K, Li Y, Wen LP (2009) *Autophagy* 5:1107
- Zhu J, Ji Z, Wang J, Sun R, Zhang X, Gao Y, Sun H, Liu Y, Wang Z, Li A, Ma J, Wang T, Jia G, Gu Y (2008) *Small* 4:1168
- Prylutska SV, Burlaka AP, Prylutskyi YI, Ritter U, Scharff P (2011) Pristine C<sub>60</sub> fullerenes inhibit the rate of tumor growth and metastasis. *Exp Oncol* 33:162–164
- Chen Z, Ma L, Liu Y, Chen C (2012) *Theranostics* 2:238
- Injac R, Radic N, Govedarica B, Perse M, Cerar A, Djordjevic A, Strukelj B (2009) *Pharmacol Reports* 61:335
- Prylutska S, Grynyuk I, Matyshevska O, Prylutskyi Y, Evstigneev M, Scharff P, Ritter U (2014) C<sub>60</sub> fullerene as synergistic agent in tumor-inhibitory doxorubicin treatment. *Drugs R D* 14:333–340
- Evstigneev MP, Buchelnikov AS, Voronin DP, Rubin YV, Belous LF, Prylutskyi YI, Ritter U (2013) Complexation of C<sub>60</sub> fullerene with aromatic drugs. *Chem Phys Chem* 14:568–578

20. Prylutskyi YI, Evstigneev MP, Pashkova IS, Wyrzykowski D, Woziwodzka A, Gołuński G, Piosik J, Cherepanov VV, Ritter U (2014) Characterization of  $C_{60}$  fullerene complexation with antibiotic doxorubicin. *Phys Chem Chem Phys* 16:23164–23172
21. Prylutskyi YI, Petrenko VI, Ivankov OI, Kyzyma OA, Bulavin LA, Litsis OO, Evstigneev MP, Cherepanov VV, Naumovets AG, Ritter U (2014) On the origin of  $C_{60}$  fullerene solubility in aqueous solution. *Langmuir* 30:3967–3970
22. Ritter U, Prylutskyi YI, Evstigneev MP, Davidenko NA, Cherepanov VV, Senenko AI, Marchenko OA, Naumovets AG (2015) Structural features of highly stable reproducible  $C_{60}$  fullerene aqueous colloid solution probed by various techniques. *Fullerenes, Nanotubes, Carbon Nanostruct* 23:530–534
23. Prylutskyi YI, Buchelnikov AS, Voronin DP, Kostjukov VV, Ritter U, Parkinson JA, Evstigneev MP (2013)  $C_{60}$  fullerene aggregation in aqueous solution. *Phys Chem Chem Phys* 15:9351–9360
24. Solyanik GI, Todor IN, Kulik GI, Chekhun VF (1999) *Exp Oncol* 21:264
25. Gharbi N, Pressac M, Hadchouel M, Szwarc H, Wilson SR, Moussa F (2005) *Nano Lett* 5:2578
26. Didenko G, Prylutska S, Kichmarenko Y, Potebnya G, Prylutskyi Y, Slobodyanik N, Ritter U, Scharff P (2013) Evaluation of the antitumor immune response to  $C_{60}$  fullerene. *Mat-wiss u Werkstofftech* 44:124–128
27. Skivka LM, Fedorchuk OG, Bezdeneznykh NO, Lykhova OO, Semesiuk NI, Kudryavets YI, Malanchuk OM (2014) *J Exp Integr Med* 4:93
28. Naumov GN, Akslen LA, Folkman J (2006) *Cell Cycle* 5:1779
29. Calderwood SK (2013) *Scientifica* 2013:217513
30. Jego G, Hazoum A, Seigneuric R, Garrido C (2013) *Cancer Lett* 332:275
31. Di Santo JP (2006) *Ann Rev Immunol* 24:257
32. Turabekova M, Rasulev B, Theodore M, Jackman J, Leszczynska D, Leszczynski J (2014) *Nanoscale* 6:3488

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